



Photosynthetic response of transgenic soybean plants, containing an *Arabidopsis* P5CR gene, during heat and drought stress

J.A. De Ronde^{a,b,*}, W.A. Cress^{c,b}, G.H.J. Krüger^d, R.J. Strasser^{d,e},
J. Van Staden^b

^aARC Roodeplaat Vegetable and Ornamental Plant Institute, Private bag X293, Pretoria 0001, South Africa

^bResearch Centre for Plant Growth and Development, University of Natal Pietermaritzburg, Private Bag X01, Scottsville 3209, South Africa

^cDepartment of Crop, Soil and Environmental Sciences, University of Arkansas, Fayetteville, AR 72701, USA

^dSchool of Environmental Sciences and Development: Botany, Potchefstroom University for CHE, Potchefstroom 2520, South Africa

^eBioenergetics Laboratory, University of Geneva, CH-1254 Jussy/Geneva, Switzerland

Received 18 June 2003; accepted 9 January 2004

KEYWORDS

Chlorophyll
fluorescence;
Drought;
Heat;
NADP⁺;
Proline;
Soybean

Summary

The biochemical basis of heat/drought tolerance was investigated by comparing the response of antisense and sense transgenic soybean plants (containing the L- Δ^1 -pyrroline-5-carboxylate reductase gene) with non-transgenic wild-type plants. The plants were subjected to a simultaneous drought and heat stress of 2 days, whereafter they were rewatered at 25°C. During this time the sense plants only showed mild symptoms of stress compared to the antisense plants which were severely stressed. Upon stress, nicotinamide adenine dinucleotide phosphate (NADP⁺) levels decreased in antisense while it increased in sense plants. Recovery with respect to NADP⁺ levels was best in sense plants. Sense plants had the highest ability to accumulate proline

Abbreviations: ABS/CS, absorption flux per leaf cross section; ABS/RC, average antenna size; CS, cross section; CF, chlorophyll-*a* fluorescence; e⁻, electron; ET, electron transport; *F*₀, maximal fluorescence intensity when all the RCs are closed; *F*₀, fluorescence intensity at 50 μ s when all RCs are open; IHSP, inducible heat shock promoter; JIP test, quantification of the Chl-*a* fluorescence *F*₀–*J*–*I*–*P* transient; NADP⁺, nicotinamide adenine dinucleotide phosphate; NADPH, nicotinamide adenine dinucleotide hydrogen phosphate; OEC, oxygen-evolving complex; P5C, L- Δ^1 -pyrroline-5-carboxylate; P5CR, L- Δ^1 -pyrroline-5-carboxylate reductase; P5CDH, P5C dehydrogenase; PDH, proline dehydrogenase; PI_(CS0), performance index expressed relative to CS; PSII, photosystem 2; RC, fully active reaction center; RC/CS₀, density of active units per leaf area; TR, trapping flux; *V*, relative variable fluorescence normalised between *F*₀ (50 μ s) and *F*_m; *V*_J, relative variable fluorescence at *J* step (2 ms); *W*, relative variable fluorescence normalised for single turn-over events between *F*₀ (50 μ s) and *F*_J (2 ms); *W*_K, relative variable fluorescence at *K* step (100–300 μ s); *Q*_A, primary quinone acceptor of PSII; ϕ (P₀), quantum efficiency of primary photochemistry (flux ratio TR₀/ABS)

*Corresponding author. ARC Roodeplaat Vegetable and Ornamental Plant Institute, Private bag X293, Pretoria 0001, South Africa. Tel.: +27-12-841-9775; fax: +27-12-808-1499.

E-mail address: kobie@igs1.agric.za (J.A. De Ronde).